

Vol. 9, Issue. 2, May 2021



Plant Secondary Metabolites: A Beneficial Resource for both Man and Butterflies-A Review

J.Naresh1V.KastroKiran2

Abstract

Traditional healers have long regarded ethnomedicine as a fascinating and appealing topic of study. This makes the proper identification of these plant species all the more vital. It's also worth noting that female butterflies are endowed with the capacity to recognise plants that are conducive to oviposition and, in turn, provide nourishment for their larvae. It is possible that butterflies' ability to choose the correct plant at the right moment demonstrates the relevance of plant secondary metabolites. This class of metabolites is also the foundation for a number of pharmaceutical compositions. As a result, this research aims to emphasise the folklore therapists' use of plant species in the tribal areas of West and East Midnapore, West Bengal, India. The identification of butterflies that feed their larvae on the same plant species was also completed.

Keywords:Butterflies and oviposition; ethnomedicine; secondary metabolites; plant secondary metabolites

1.Introduction

Nature has been man's closest friend since the start of civilisation. Even in times of crisis, Nature has been there to help those in need. The ancient systems of medicine (such as Ayurveda, Unani, Siddha, and Homeopathy) remain very popular in spite of the fast urbanisation and contemporary technical advances. Considering that the Rig Vedas reference herbal treatments, and that Indian herbalists such as Maharishi Charaka and Sushruta [1] used them, it's interesting noting

how widespread their use has been. Significantly, the WHO has found that 80 percent of individuals in underdeveloped countries choose traditional treatments. [2] Around 8000 kinds of medicinal plants have been discovered among the approximately 17,500 flowering plant species that have been identified as being native to India. [3]

Assistant Professor, Department of Civil Engineering, Bharat Institute of Engineering and Technology, Telangana, India¹

 $Assistant Professor, Department of Civil Engineering, Vignan Institute of Technology and Science, Telangana, India^2$

For forest fringe people, the abundance of nature's resources seems to be a source of ethnomedicine. The rising demand for these plants may be attributed to the widespread usage of traditional herbal remedies, particularly among rural populations. Folklore therapy's use of herbal formulations has become so widespread that effective resource identification and usage is essential.

Insects, on the other hand, are as efficient and possibly even more thorough in their selection of host plants. A wide variety of insect species have evolved a variety of physiological adaptations in order to overcome the plant taxa's defence substrates (i.e., plant secondary metabolic compounds). Four, five, and six] There is an intimate interaction between butterflies and their habitats because of their need on flora, both as adults and as larvae. Nectar resources are required as they mature. The "Botanical Instinct" of butterflies may be shown in their tendency to choose plants that are closely linked to one other for egg depositing. [7] In their larval stages, butterflies often show host plant preference by being either single- or multiple-feeders.

As a result, the larval plant interaction is heavily influenced by their selection of larval host plants. Species-specific secondary metabolites of plants are critical in the selection of larval feeding sources. [8] Additionally, a wide variety of plant secondary metabolites, such as aromatic compounds, nitrogen-containing metabolites like alkaloids, cyanogenic glycosides, glucosinolates, and other isopyrenoids, such as cardiac glycosides, cucurbitacins, iridoid glycosides, and others, are extremely popular among medical practitioners. [9]

Plant Secondary Metabolites (PSMs) play a critical role in the lives of butterflies, and they also serve as the foundation for many folkloric herbal formulas.

2. StudyArea

Surveys were undertaken in West Bengal's East Midnapore and West Midnapore, two tribal strongholds. Ethnomedicine is a growing topic in West Bengal, and this research aims to take use of the knowledge of people who live in the region's main tribal areas. Despite their strong religious and cultural connections, their poverty and lack of access to modern medical care have resulted in an increasing reliance on herbal remedies. Herbal compositions based on such in-depth understanding of medicinal plants have given rise to various modern-day medications. [12]

3. **Methodology**

4.Based on informal face-to-face interaction, ethnomedicinal surveys were done with the local population of all castes and faiths with the hope of identifying the most informed individuals (old, aged and experienced people, ojhas, kaviraj and vaidyas). Using a series of semi-structured open ended questions that had been developed in advance, we interviewed a total of 60 informants. In addition to group discussions and casual meetups, if needed, These folk therapists passed on their expertise of folk medicine in a data sheet. No changes were made to the depiction of information supplied by informants, which is notable. These participants' hypotheses about the therapeutic potential of some plants were eventually corroborated by peer-reviewed publications. [3] Photographs of several plant specimens were taken, and the taxonomist's help was used to identify them [13, 14, 15].

5.In the last stage of our research, we looked for butterfly species that feed their larvae on the same plants. Importantly, available literature was used to help characterise larval host plants. [16,17]

6. Secondary Metabolic constituents of ethnomedic in alplants pecies

A list of plant secondary metabolites (i.e., alkaloids, terpenoids, glycosides, flavonoids, tannins, cardenolides, and sterols) previously

documented from various ethnomedicinal plant species is provided in this paper.

Table1.

Table 1. List of major phytochemical constituents found in plant species

S1.	Name	Major phytochemical constituents		
No.				
1	Hygrophila auriculata Schumach.	Alkaloids: asteracanthicine, asteracanthine Flavonoids: apigenin 7-O-glucuronide, apigenin-7-O-glucoside Terpenoids: lupeol, betulin, luteolin, luteolin-7-O-retinosides [18]		
2	Aristolochia indica L.	Terpenoids: α-pinene, pinocarvone, trans-pinocarvol [19]		
3	Calotropis gigantea (Linn.)Aton f.	Cardiac glycosides: calotropogenin, calotropin [20] Terpenoids: α-amyrin, β-amyrin, α-amyrin methylbutazone, β-amyrin methylbutazone, α-amyrin acetate, β-amyrin acetate, lupeol acetate. [21]		
4	Cryptolepis buchanani Roem. & Schult.	Alkaloids: buchananine Glycosides: sarverogenin, isosarverogenin Cardenolides: cryptosin, buchanin [22]		
5	Tylophora indica (Brum. f.) Merr.	Alkaloids: tyloindicines A-J, phenanthroindolizidine alkaloids (tylophorine, tylophorinine, tylophorinidine Flavonoids: quercetin, kaempferol Terpenoids: α-amyrin, β-amyrin [23]		
6	Capparis zeylanica L.	Glycosides: saponin glycosides Terpenoids: α-amyrin, β-carotene [24]		
7	Quisqualis indica L.	Tannins: Ellagitannins quisqualin-A, quisqualin-B, [25], gallic acid [26] Flavonoids: quercetin, rutin [27, 26] Terpenoids: β-sitosterol, lupeol [26]		
8	Ricinus communis L.	Alkaloids: ricinine Flavonoids: rutinoside, kaempferol derivatives Tepenoids: 1-8 cineole, camphor, α-pinene, β-caryophyllene, epicatechin [28]		
9	Albizia amara (Roxb.) B. Boivin	Alkaloids: budmunchiamines [29]		
10	Cassia fistula L.	Glycosides: sennoside-A, sennoside-B Flavonoids: epiafzelechin, epicatechin, biflavonoids, procyanidin B, triflavonoids [30]		
11	Cassia tora L.	Flavonoids: quercitin, iso-quercitin, emodin, friendlein [31]		
12	Dalbergia sissoo Roxb. ex DC	Flavonoids: dalbergichromene, dalbergin, dalberginone, methyl dalbergin [32]		
13	Mimosa pudica L.	Alkaloids: L-mimosine. pinitol Flavonoids: 5 deoxyflavonol derivatives, kaempferol-3-rutinoside, leutolin-3-xyloside, acaetin-7-rutinoside Glycosides: quercetin-7-rhamnoside, quercetin 3-glucoside 7-rhamnoside [33]		
14	Tamarindus indicus L.	Terpenoids: limonene. caryophyllene, lupanone, lupeol, p-cymene, longifolene [34]		
15	Cinnamomum camphora (L.)J. Presl.	Terpenoids: cineol, limonene, α-humulene, β-cardinine, D-camphor, α-terpineol, linalool, γ -terpinene, α-pinene, β-pinene, eugenol [35, 36, 37]		
16	Ficus glomerata Roxb.	Terpenoids:Lanosterol, gluanol acetate, lupeol, α-amyrin acetate, lupeol acetate. Sterols: stigmasterol [38]		

17	Streblus asper Lour.	Terpenoids: Lupeol, α-amyrin
		Sterol: β-sitosterol [39]
18	Oxalis corniculata L.	Flavonoids: Isoorientin, isovitexin, sertisin,
		Terpenoids : β-sitosterol, betulin [40]
19	Cymnopogan citratus	Terpenoids: citral (geranial and neral) [41]
	(DC.) Stapf	
20 Portulaca oleracea L. Glycosides: Cardiac		Glycosides: Cardiac glycosides, anthraquinone glycosides
		Flavonoids: kaempferol, quercetin, luteolin, myricetin, genistin
		[42]
21	Zizyphus oenopalia L.	Alkaloids: Zizyphine (A, B, C, D, E, F, I, K, N, O, P), frangufoline,
	(Mill.)	mauritine-D, abyssinine B, A; amphibine B, F [43]
22 Aegle marmelos L. Alkaloids: Aeglin, aegelenine. dictam		Alkaloids: Aeglin, aegelenine. dictamine, fragrine [44]
		Terpenoids :α-phellandrene, P-cymene; limonene [45]
23	Murraya koenigi(L.)	Alkaloids: koenine, koenigine, girininbine, koenimbine, isomahanine,
	Spreng.	bismahanine. Coumarin glycosides : scopotin, murrayanine [46, 47]
24	Grewia asiatica L.	Flavonoids :quercetin, kaempferol, Terpenoid : taraxerol, erythrodiol,
		lupeol, lupenone, friedlin,β-amyrin [48]
25	Cinnamomum tamala	Terpenoids :Phellandrene, eugenol, linalool, α-pinene,
	(Buch. – Ham.)T. Nees	p-cymene, β-pinene, limonene, phenyl propanoids, furanogermenone [49]
	& C. H. Eberm.	
26	Plumbago zeylanica L.	Plumbagin [50]

7. Utilization of plants econdary metabolites by larval stages of butterflies: Glycosides of the cardiovascular system.

A class of insects specialised in sequestering cardiac glycosides from food sources has been shown to improve insect defences. [51,52] Butterflies that feed on cardiac glycoside-rich Asclepius plants, such as Danainae, are able to integrate these compounds into wing and integument structures. [54,55] The capacity of specialised insects to feed on plant-derived glycosides has been carefully studied. 51 and 52 Insects that have developed the ability to feed on chemically enhanced plant tissues have likely contributed to the diversification of many insect species. Sixty-six to sixtieth.

Flavonoids

Some larval lycaenids store flavonoids from their host plants for integration into their wings, which is the foundation of insect-plant interaction. 61 and 62 Adult butterflies' flavonoid content is closely linked to the percentage of their larval host plant that they ingest. 62, 63, and 64 Flavonoids (especially isovitexin) are being metabolised by larval Pseudozizeeriamaha when it is eating on Oxalis corniculata. Both of these statements are true [65–66]. As a result, the abundance of isovitexin

7-O-glucoside in their larval stages was converted into isovitexin in the pupae. [66] The inflorescence of Medicago sativa is stated to be richer in flavonoid as compared to the solitary leaf of the same plant when fed to lycaenid larvae. [67] It is well-known that herb-feeding lycaenids are capable of manipulating the quantitative defence chemicals present in Fabales, Santalales, and Rosales with great success. When you're 68 and 69 years old, Aristolochiaceae, Apocyanaceae, Asclepiadaceae, and Brassicaceae are known to avoid defence substances like tannins and polyphenolics, which are prevalent in woody plants. [p. 68, p. 69] Caterpillars' ability to resist plant defences by removing harmful chemicals is one of the factors contributing to their unusually wide diet. [70] UV radiation is absorbed by flavonoids, resulting in a brighter, more vibrant colour. [71] Women may store 60 percent more flavonoids than men do, making this a significant difference in health. Flavonoid content in the form of UV patterns on their wings indicates a higher level of female quality and an improved ability to recognise potential mates.

Alkaloids:

Aristolochic Acids (AA), notably AA-I and AA-II, make Aristolochiaceae distinctive in terms of their pharmacological properties. After this, AA-I and AA-II are said to be converted metabolically, whereas AA-II is said to be converted selectively. [72] Larvae of Troidini were able to extract Aristolochic Acids and benzylisoquinoline from their host plants, which they then deposited in their bodies. 73.3 and 74.4 The fact that these butterflies have achieved such a high level of sequestration renders them unappetizing, dangerous, and poisonous to their predators. However, the

osmeterial glands of these larvae secrete aristolochic acids when agitated. [52] Mullerian convergence in combination with apostolic colouring gives them ideal examples of mimicry. [75] It's remarkable that Papilionidae use osmeterium for defence reasons in this way. [52,76]

8. PlantSecondaryMetabolitesabeneficialresourceforbothmanandbutterflies Table 2 lists human reports of the medical effects of the plant species. Intriguingly, the same plants are used as ovipositing substrate by female butterflies at this research location as well. Table 2.

Table2.Listofplantspeciesbeneficialforbothmanandbutterflies

S1.	Name	Habitat	Medicinal benefits as reported	Ovipositing
No.			among humans	butterfly species
1	Hygrophila auriculata Schumach.	marshlands	Possess antitumor, antibacterial, antioxidant, hepatoprotective properties [18]	Junonia orithiya, J.lemonias, J.atlites,J.iphita, J. hierta
2	Aristolochia indica L.	Stream bank, degraded land	Effective against cholera, fever, bowel problems, ulcers, leprosy, snake bites and skin diseases. [77] Also being employed as an abortifacient, antineoplastic [77]	Atrophaneura hector, A. aristolochiae
3	Calotropis gigantea (Linn.) Aton f.	stream bank, roadsides	Pharmacological activities includes it's use as an antiseptic, anti inflammatory, hepatoprotective, antimyocordial infraction activity and anti diarrhoeal. Plant also beneficial for the treatment of leprosy, tumor and ulcers. [78]	Danaus chrysippus, Parantica aglea
4	Cryptolepis buchanani Roem. & Schult.	along jungle edges	Used in the treatment of rickets. Decoction of leaves when mixed with honey cures dropsy. Paste of leaves, stem and roots are also applied on ulcers. [3]	Parantica aglea Euploea core
5	Tylophora indica (Brum. f.) Merr.	wasteland	Possess immunomodulatory, antiimflammatory and anticancer properties [79, 80]	Tirumala limniace, T. septentrionis, Danaus genutia, D. chrysippus, Parantica aglea
6	Capparis zeylanica L.	hedges	Used as counter irritant and febrifuge. Also effective against cholera and swelling of testicle, boils and small pox. [81]	Hebomoia glaucippe, Appias libythea, Leptosia nina

7	Quisqualis	disturbed	Beneficial against flatulence.	Rapala manea,
	indica L.	area	Decoction prepared from leaves used	R. varuna
			to treat asthma and intestinal worms	
			[3]	
8	Ricinus	wasteland	Useful in the treatment of night	Ariadne ariadne
	communis		blindness, opthalmia, headache,	A. merione
	Linn.		earache, corpulence and cough [3]	
9	Albizia amara	rocky edges	Provides relief from piles, diarrohea,	Charaxes bernardus,
	(Roxb.) B.		gonorrhea, leprosy, leucoderma,[29]	Eurema hecabe
	Boivin		and opthalmia [3]	
10	Cassia fistula	low hill sides	Possess antipyretic, analgesic, [82]	Eurema brigitta,
	L.		anti-tussive and wound healing [83,	E. hecabe, Catopsilia
			84] properties	pomona, C.pyranthe
11	Cassia tora	wasteland	Cures conjunctional congestion caused	E. hecabe, atopsilia
	Linn.		by liver fire, blurring vision and	pomona, C.pyranthe
			constipation. Also found to be	
			antifungal, anti inflammatory and	
			antihepatotoxic in nature [31]	
12	Dalbergia	riverbanks	Known to exhibit anti microbial,	Neptis hylas,
	sissoo: Roxb.		cardiac, antioxidant, antiparasitic,	Phaedyma columella,
	ex DC		antidiabetic, anti-inflammatory,	Charaxes bernardus
			analgesic, osteogenic, dermatological,	
			gasterointestinal and reproductive	
			effects [32]	
13	Mimosa pudica	weed,	Widely recognised for it's	Prosotas nora,
	L.	croplands or	antinociceptive, antihelmintic,	P. dubiosa, Junonia
		pastures	hyperglycaemic, immunomodulatory,	orithiya
			anticonvulsant, diuretic and wound	
			healing properties [85, 86 87, 88, 89]	
14	Tamarindus	wild or	Leaf juice used to treat bleeding piles,	Charaxes bernardus,
	indicus L.	disturbed	painful anuria, pox, dysentery, ulcer	C. solon
		area	and rheumatic pain. Stem used as	
			antipyretic and astringent [3\	
15	Cinnamomum	Roadsides,	Camphor used to treat high fever,	Graphium sarpedon,
	camphora (L.)	forest edges	measles, delirium, whooping cough,	G. epycides, Chilasa
	J. Presl.	or woodland	melancholia, chronic bronchitis.	clytia

19	Cymbopogon	disturbed	Leaf oil employed to provide remedy	Parnara guttatus,
	citratus Stapf.	areas	against flatulence, spasmodic affection	Ampitta dioscorides
			of bowel, gastric irritability, cholera,	
			chronic rheumatism and sprains [3]	
20	Portulaca	disturbed	Acts as an antiseptic.	Hypolimnas bolina,
	oleracea L.	area,	antispasmodic, vermifuge,	
			antiscorbutic, antibacterial, analgesic,	
			anti-inflammatory, skeletal muscle	
			relaxant, bronchodilator, antipyretic	
			and antiasthma [42]	
21	Zizyphus	roadside	Widely recognised febrifuge and tonic.	Tarucus nara,
	oenopalia L.		Effective in treatment of inflammation	
	(Mill.)		of mucus lining of uterus and to cure	
			dysentery [3]	
22	Aegle	cultivated dry	Known to lower blood glucose levels.	Papilio polytes, P.
	marmelos	forest	[90] Plant possesses antifungal [91]	demoleus
	Correa.		and antispermatogenic effects. [92]	
23	Murraya	cultivated	A number of pharmacological	Papilio polytes, P.
	koenigi (L.)	land	activities such as anti-tumor, anti-	demoleus
	Spreng.		oxidative, anti-mutagenic and anti-	
			inflammatory have been reported [93,	
			94]	
24	Grewia	scrubland	Widely recognised as an effective	Neptis jumbah, N.
	asiatica L.		febrifuge, demulcent, Also possess anti	hylas
			diarrohea [48] properties	
25	Cinnamomum	riverbed	Is known to be an astringent with	Graphium
	tamala (Buch.		carminatine properties. Beneficial for	agamemnon
	– Ham.)T.		the treatment of itch, diarrhoea, colic,	
	Nees & C. H.		rheumatism, fever, anaemia, nausea	
	Eberm.		and vomiting [49]	
26	Plumbago	riverbed	Paste prepared from stem bark	Leptotes plinius
	zeylanica L.		effective against piles, Leaf paste	
			applied to swelling and elephantiasis.	
			[3]	

5. **Concludingremarks**

Due to their need to survive and thrive at various points in their lives, butterfly larvae may have developed an intimate connection with a small number of plants that produce plant secondary metabolites. Butterflies have been

reported to stow away unappetizing or harmful metabolites in order to protect themselves from predators. Aspirin, digitalis, and certain anti-cancer and anti-malarial medications have all come from plant origins, despite their

widespread usage in contemporary medicine (Grover et al 2002). As a result, ethnobotany may be used to develop new herbal compositions by drawing on the traditional knowledge of the ethnic group's members.

Ethnomedicinal plants that butterflies consume as larvae urge for careful use of these resources by humans.

The already fragile ecology of India's Midnapore districts has been further weakened by a lack of scientific skills and information, as well as by growing urbanisation. [10] From this viewpoint, the current research in East and West Midnaporeis noteworthy.

6. Acknowledgement

Thank you to Professor Narayan Ghorai of the Zoology Department of West Bengal State University, India for your important advice and assistance over the course of this research project. Thank you to everyone who participated in the research and shared their wisdom and insights with us, especially the informants!

References

Several ethno-medicinal plants of the Lamiaceae family have been documented in Bankura district, West Bengal, India, according to a study by A. Sinhababu and A. Banerjee.

Ethnomedicinal plants of West Bengal's West Rarrh area have been documented in Natural Product Radiance, a journal devoted to the study of natural products, with an issue number of 7.

[3]Natural Resources Directorate of the West Bengal Government, "Medicinal Plant Resources of South West Bengal," (2005) xi + 198Noel Theis and Michael Ledau "The Evolution of Secondary Metabolites' Functions," International Journal of Plant Sciences, 164 (2003) S93-S102. [4]

An ecological and molecular evolutionary viewpoint on secondary metabolites has been presented by M. Wink in Phytochemistry, vol. 64, no. 1 (2003).

When it comes to insect herbivores, it's important to know the difference between "specialised" and "generalist" insects.[7] S. V. Ramana, "Biodiversity and conservation of butterflies in the Eastern Ghats," The Ecoscan, Vol. P4, No. 1, (2010), pp. 59-67.

Ecology of Butterflies in Europe, Cambridge University Press (Cambridge, 2009), pp: xii+513, by J. Settele, T. Shreeve, M. Konivika, and H. Van Dyck.

Medical herbs of India having antidiabetic potential, Journal of Ethnopharmacology, volume 81, number 2, 2002, pp. 81-100.

Traditional folk medicine practises of the Kora-Mudi tribe of PaschimMedinpur, West Bengal, have been documented by researchers [10] P Halder, S Mukherjee and M Bhattacharya. National Conference on Traditional Knowledge and Social Practice, 2014, pgs.101-108.

Ethnogynaecological applications of plants by the Lodha people in PaschimMedinipur district, West Bengal, are documented in the World Journal of Alternative Medicine, volume 1, issue 1, pp. 1-4, in 2014. [11]

"Traditional usage of medicinal herbs as febrifuges by the tribals of Purulia district, West Bengal, India," Asian Pacific Journal of Tropical Disease, pp. S800-S803 (2012).In O. Polunin and A. Stainton (eds.), Flowers of the Himalaya, Seventh Edition Oxford University Press, New Delhi (2005), pp. xxx+580.

(2001) pg x+430 in "Annotated checklist of Nepal's blooming plants" by J. R. Press, K KShrestha and D A Sutton at the Natural History Museum in London (2001).

N. P. Manandhar, "Plants and people of Nepal," Timber Press, Inc. Portland, USA (2002), pages 599.

16] "The Butterflies of Sikkim Himalaya and Their Natural History," Sikkim. Conservation Foundation of Sikkim Natural Resources.Gangtok, (1992), pp. 217.

Indian Butterflies by I. Kehimkar, Bombay Natural History Society. [17] New Delhi, 2008: Oxford University Press.

Preliminary research on the diuretic activity of Hygrophilaauriculata (Schum) 'Heine in rats

were conducted by Md. S. Hussain, K. F. H. N. Ahmed, and Md. Z H Ansari.

An essential oil stem oil from Aristolochiaindica L. has been studied for its chemical composition by P. B. Kanjilal, R. Kotoky, and M. Couladis.

This research was published in Science in 1964 in the pages 1685-1686 as "Calotropin, cytotoxic principle extracted from the Asclepiascurassavica L." [20].

Flavonol glycosides from Calotropisgigantea were discovered by Sen, N. P. Sahu, and S. B. Mahato in 1992."Development of Pharmacognostical profile of CryptolepisbuchananiRoem&Schult" was published in International Journal of Pharmacy and Pharmaceutical Sciences, Vol. 4, No. 2, (2012), pp. 615-618.

There is a lot of information out there on the medicinal properties of Tylophoraindica (Burm f.) Merill, but this article provides a comprehensive analysis of the plant's pharmacological and plant tissue culture properties.

(IJRAP): "Phytochemistry and pharmacological activity of Cappariszeylanica: An overview," IJRAP (IJRAP, vol. 1, no. 2, 2010): 384-389.

"Tannins and Related Compounds from Quisqualisindica" was published in the Journal of the Chinese Chemical Society, Vol. 44, 1997, pp. 151-155.

V. A. Bairagi, P. R. Shinde, K. L. Senthikumar, and N. Sandu, "Isolation and characterizations of phytoconstituents from the Quisqualisindica Linn. (Combretaceae)," Research Journal of Pharmacognosy and Phytochemistry, vol. 4, no. 4, (2012), pp 229-233.

[27] "Flavonoids of the leaves and flowers of Quisqualisindica Linn" by G. A. Nair, C. P. Joshua, and A. G. R. Nair. Vol. 18, No. 3, (1979), pp: 291-292 in Indian J. Chem B

IJPPS International Journal of Pharmacy and Pharmaceutical Sciences, 4, no. 4, (2012), pp: 25-29: "RicinuscommunisLinn.: A Phytopharmacological Review."

"Chromatographic fingerprint study of budmunchiamines in Albiziaamara by HPTLC

approach," Int. J. of Res. Pharm. Sci., vol. 1, no. 3, pp. 313-316, 2010.

[30] "Phytochemical components of Cassia fistula," by T. Bahorun, V. S. Neergheen, and O. I. Aruoma. Volume 4, Number 13, pp. 1530–1540, African Journal of Biotechnology, (2005). According to the Indian Journal of Natural Products, Cassia tora Linn.has been studied extensively in terms of its pharmacological and phytochemical properties.

This article, "Chemical components and pharmacological effects of the Dalbergiasissoo - A Review," was published in the Journal of Pharmacy, vol. 7, no. 2, (2017), pp59–71.

According to the authors, "Mimosa- A quick summary" was published in Journal of Pharmacognosy and Phytochemistry, Volume 4, Number 2, in 2015.

"Chemical components of Tamarindusindica L. leaves" was published in RevistaCubana de Quimica, volume 22, issue 3, pp.65-71 in 2010.

[15] Ethnobotanical Leaflets, Vol. 12, pp. 181-190, (2008).

[16] For example, [17] C-L. Ho, I-C. Wang Eugene and Y-C. S wrote in LinalooliferaFujuta that "Essential oil compositions and bioactivities of the different sections of Cin

[17]The namomumcamphoraSieb var." An article published in Chemistry and Biodiversity, volume 6, no. 9, (2009), entitled "Terpenoid diversity in the leaf essential oils of Himalayan Lauraceae species" provides an overview of the terpenoid composition of these oils.

[18]"Ficusracemosa: Phytochemistry, traditional applications, and pharmacological qualities – A review" by A. R. Shiksharthi and S. Mittal. Pharmaceutical Research, vol. 4, pages. 6-15, 2011.

[19]Streblusasper (Shakhotaka): a review of its chemical, pharmacological, and ethnomedicinal qualities. Evidence-based Complementary and Alternative Medicine, 3, no. 2, 2006, pp. 217-222;

[20]A review of the phytochemistry and pharnacology of Oxalis coniculata Linn. – A review of the pharnacology and pharnacology of Oxalis coniculata Linn. – A review

[21]"Histochemical location of citral buildup in lemongrass leaves (Cymbopogoncitratus (DC.) Stapf.,Poaceae)".

[22]Studies on Portulacaoleracea (purslane) Phytochemistry by I. AOkafor and D N Ezejindu (23). pp. 132-136 in the Global Journal of Biology, Agriculture, and Health Sciences.

[23]In vitro anti-denaturation and antibacterial activity of Zizyphusoenoplia have been studied by R. Ramalingam, B BMadhavi, A. R Nath, N Duganath, E. U. Sri, and D. Banji.

[24]New alkaloids from Aeglemarmelos have been discovered by M D Manandhar, A Shoeb, R S Kapil, and S P Popli in Phytochemistry, vol. 17, (1978), pp. 1814-1815.

[25]Leaves of Aeglemarmelos Linn, "Phytochemical and pharmacological characteristics", The Pharmaceutical Reviews, (2009), pp.144-159, N. P. Yadav and C. S. Chanotia

[26]YA comparison of the antioxidative effects of carbazole alkaloids from Murrayakoenigii leaves by Tachibana, Kikuzaki, Lajis, and Nakatani, J. Agric Food Chemistry, vol. 51 (2003), pp. 6461-6467

[27]Methanolic extract and certain carbazole alkaloids extracted from the leaves of Murrayakoenigii growing in Nigeria were shown to have anti-trichomonal, biochemical, and toxicological properties, according to the study published in Phytomedicine, vol. 13, pp. 246-254 in 2006.

[28] Pharmacological characteristics of Grewia are reviewed in International Journal of Pharmacy and Pharmaceutical Sciences, volume 4, no. 4, (2012), pages 72-78, by P. K. Goyal. [29] [29]

[29]IJPSRR: International Journal of Pharmaceutical Sciences: Review and Research, Vol. 5, No. 3, 2010, pp.141-144. Cinnamomumtamala'sethnopharmacological properties: A review

[30]Plumbagozeylanica Linn. aerial portions have been shown to have hepatoprotective efficacy against carbon tetrachloride-induced hepatotoxicity in rats, according to a study published in the International Journal of

Pharmacy and Pharmaceutical Sciences, volume 1, number 1, 2009, pages 171-175.

[31]Dobler, D. Daloze and J. M. Pasteels, "Cardenolides in Chrysochus" in Chemoecology, vol. 8, pp111-118, (1998), cited in this article

[32]A study published in the journal Annual Review of Entomology in 2002 found that lepidopteran species sequester defence chemicals from plants.

[33]M. Aardema is a Molecular Ecology, vol. 21, no. 2, 2012, pp. 340-349, Y. Zhen and P. Andolfatto, "The development of cardenolideresistant variants of Na+, K+- ATPase in danainae butterflies."

[35] Frick and Wink [35] Ouabain uptake and sequestration in Danausplexippus (Lepidoptera: Daninae): evidence for carrier-mediated mechanism, Journal of Chemical Ecology

[37]Ecology, Vol. 21, pp. 557-575, 1995.

[38]Studies on the sequestration of cardenolides in African milkweed butterflies (Danaidae) have been published in Toxicon, volume 45, issue 5, pp. 581–584.

[39]P.R. Ehrlich and P.H. Raven, "Butterflies and plants: a study in coevolution," Evolution, vol.18, (1964), p. 586-608. [40]

[40]]Cytochrome P450s are two examples of "important innovations" in an adaptive radiation, according to Berenbaum, Favret, and Schuler in the American Naturalist.

[42]In "Both host-plant phylogeny and chemistry have affected the African seed-beetle radiation," Molecular Phylogenetics and Evolution, vol. 35, no. 3, (2005), pp. 602–611, G.K. Kergoat, A. Delobel, G.Fédière and B Rü, J.F. Silvain.

[44]"The genetic foundation of a plant-insect coevolutionary major innovation," Proceedings of the National Academy of Sciences, 104 (2007), 20427-20431. S

[45]Recently, we've learned a lot more about how insects and flavonoids interact. Phytochemistry, 64(1), pp. 21-30, 2003.

[46]Journal of Chemical Ecology, Vol. 13, 1987, pp. 473-493, by A. Wilson, "Flavonoid pigments in the chalkhill blue (LysandracoridonPoda) and other lycaenid butterflies."

[47]Journal of Chemical Ecology, vol. 20, (1994), pp. 2523-2538. "Sequestration of host plant-derived flavonoids by lycaenid butterfly Polyommatusicarus."

[48]In the case of Polyommatusicarus (Lepidoptera:Lycaenidae), the lycaenid butterfly, the uptake of flavonoids from Viciavillosa (Fabaceae) by Polyommatusicarus (Lepidoptera:Lycaenidae) was studied.

[49]Flavonoids are found in the common blue butterfly Polyommatusicarus if it is grown on Trifoliumrepens, according to research published in Phytochemistry in 1999 by U. Schittko and his colleagues.

[50] H. Mizokami, K. Tomita-Yokotani, and K. Yoshitama, "Flavonoids in the leaves of Oxalis corniculata and the sequestration of the flavonoids in the wing scales of the light grass blue butterfly, Pseudozizeeriamaha," Journal of Plant Research, vol. 121, no. 1, (2008), pp. 133-136. [51].

[51] H. Mizokami and K. Yoshitama, "The Pale Grass Blue, Pseudozizeeriamaha (Lepidoptera: Lycaenidae)," Entomological Science, vol. 12, pp. 171-176, (2009)

Female common blue (Polyommatusicarus) butterflies with flavonoid wing pigments are more appealing to mate-hunting males, according to a study published in the journal Naturwissenschaften.

In Recent Advances in Phytochemistry, vol. 10, (1976), pp.1-40, P. P. Feeny discusses "plant appearance and chemical defence."

[52]D. F. Rhoades and R. G. Cates, "Towards a general theory of plant anti herbivore chemistry",

RecentadvancesinPhytochemistry,vol.10,(1976), pp.168-213.

[53]K.Fiedler, "Host-plantrelationshipsof lycaenid butterflies: largescalepatterns, interactions with plant chemistry, and mutualism with ants'. Entomologia Experimentaliset Applicanta, vol. 80, (1996), pp.259-267.

[54] K Lunau"A new interpretation of flower guide colours: absorption of ultraviolet light

enhances coloursaturation",Plant Syst.Evol.,vol.183,(1992),pp.51-65.

[55] K.R. Sime, P.P. Feenyand M. Haribal, "Sequestr ation of a ristolochicacids by the pipe vines wallow ta il, Battus philen or (L.): evidence and ecological implications", Chemoecology, vol. 10, (2000), pp. 169-178.

[56] K.S. Brown, J.R. Trigo, R.B. Francini, A.B.B. Morai s, and P.C. Motta.. "Aposematic insects on toxic host plants coevolution, colonization and chemical emancipation, Plant-

AnimalInteractions", Evolutionary Ecology in Tropical and Temperate Regions, (1991), pp. 375-402.

[57]A. Urzûa and H. Priestap, "Aristolochic acids from Battuspolydamos", Biochemical Systematics and Ecology, vol. 13, (1985), pp. 169-170.

[58]A.B.B.deMorais, and K.S.Brown Jr, "Larval food plantand other effects on troidine guild composition (Papilionidae) in southeastern Brazil", The Journal of Research on the Lepidoptera, vol. 30, (1992), pp. 19-37.

[59]J.Chattopadhyay."Thestructureanddefensiv eefficacyofglandularsecretionofthelarvalosmete riuminGraphiumagamemnonagamemnonLinnae us,1758 (Lepidoptera: Papilionidae)",TurkishJournalofZoology,vol.35,n o.2,(2011),pp.245-254.

[60]A.DeyandJ.N.De, "AristolochiaindicaL.:AReview", Asian Journal of Plant Sciences, vol. 10, no. 2, (2011), pp. 108-116.

[61]A.Joshi, N.Singh, A.K.Pathakand M.Tailang, "Phytochemistryandevaluation of antioxidant activity of whole plant of Calotropisgigantea Linn.", International Journal of Research in Ayurveda and Pharmacy, vol. 1, no. 1, (2010), pp. 120-125.

[62]C.Gopalkrishan, D.Shankarnaryanan S.KmNaz imudeen, LKameshwaran, "Studies of pharmacolo gical effects of extracts and total alkaloids of Tylophoraindica", Indian J. med. Res, vol. 71, (1980), pp. 940-948.

[63]T,Ganguly,L.P.BadhekaandK.B.Sainis,"Immu nomodulatoryeffectonConAinducedlymphoproli feration", Phytomedicine, Proquest Health and Medical complete, vol. 8, no. 6, (2001), pp.431-437.

[64] K.R. Sini, B.N. Sinhaand A. Rajasekaran, "Protective Effects of Cappariszeylanica Linn. leaf extracton gastriclesions

inexperimentalanimals", Avicenna Journal of Medical Biotechnology, vol.

3,no.1,(2011),pp.31-35.

[65]D. Patel, D. Karbhari, D. Gulati and D. Gokhale. "Antipyretic and analgesic activities of AconatumspicatumandCassiafistula", Pharmace uticalBiology, vol. 157, (1965), pp. 22-27.

[66]T. Bhakta, P. K. Mukherjee, M. Pal and B. P. Saha, "Studies on antitussive activity of Cassia fistula(Leguminosae)leafextract", Pharmaceutica lBiology, vol. 36, (1998a), pp. 140-143.

[67]T. Bhakta, P. K Mukherjee, K. Mukherjee, M. Pal and B. P. Saha, "Studies on in vivo wound healingactivity of Cassia fistula Linn. leaves (Leguminosae) in rats", Natural Product Sciences, vol. 4, (1998b), pp.84-87.

[68]M. Karthikeyanand M. K. Deepa, "Antinociceptive activity of Mimosa pudica Linn", Iranian Journal of Pharmacology & Therapeutics, vol. 9, no. 1, (2010), pp. 11.

[69]R.D. Bendgude, M. G. Maniyar, M. S. Kondawar, S. B Patil, and R. V.Hirave, "Anthelminthicactivity of leaves of Mimo sapudica", International Journal of Institutional Pharmacy and Life Sciences, vol. 2, no. 1, (2012), pp. 120-123.

[70]A. Baghel, D. S. Rathore and V. Gupta, "Evaluation of diuretic activity of different extracts of MimosapudicaLinn.", PakistanjournalofBiological Sciences, vol. 16, no. 20, (2013), pp. 1223-1225.

[71]N. Alasyam, M. Sunil, T. Jayasree, V. K. Kumar, C. Nagesh and N. Venkatanarayana, "Evaluation ofanticonvulsant activity of alcoholic extract of Mimosa pudica in swiss albino rats", Journal of Chemical and Pharmaceutical Research, vol. 6, (2014), pp. 11 75-1179.

[72] G. Venkateshwarlu, K. Vijayabhaskar, G. Pavan kumar, P. Kirankumar, K. Harishbabu, and R. Maloth u, "Woundhealing activity of Mimosapudicainal bi now istarrats", Journal of Chemical and Pharmaceutical Research, vol. 3, no. 5, (2011), pp. 56-60.

[73]The Ayurvedic Pharmacopoeia of India, Part-I vol. -IV, 1stedn., Govt. of India. Ministry of Health andFamilyWelfare,DeptofAYUSH,NewDelhi,(198 8),pp.xii+167.

[74] Renu. "Fungitoxicity of leaf extracts of some higher plants against Rhizoctoniasolani Kuehn.", Nat. Acad. Sci. Lett, vol. 6, (1983), pp. 245-246.

[75]T.K.Sur,S.PanditandT.Pramanik,"Antisperma togenicactivityofleavesofAeglemarmelosCorr.in albinorats:apreliminaryreport",Biomed,vol.19,(1999),pp.199-202.

]93]K.B.Chakravarthy,S.GupthaandK.Tode,"Functionalbeta-

cellregenerationintheisletsofpancreasinalloxan-induceddiabeticratsby(-)-

epicatechin",LifeSciences,vol.31,(1982),pp.2693 -2697.

[94] P. Muthumani, V. K. Ramseshu, R. Meeraand P. Devi, "Phytochemical investigation and antimicrobial and enzymeinhibition activity of Murraya koenigii (Linn).", International Journal of Pharmace utical and Biological Archives, vol. 1, (2010), pp. 345-349.